

Deep CRISPR Nuclease Portfolio and Multiple Editing Modalities Accelerates Identification of Viable Clinical Candidates

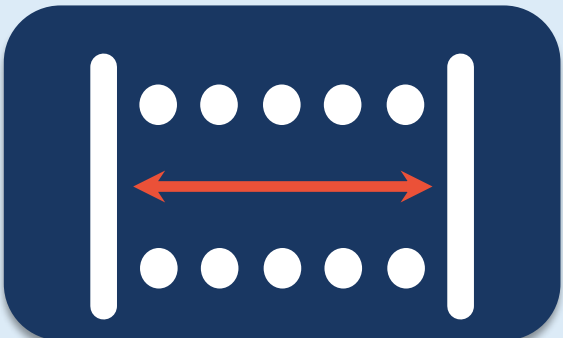
Vasu Kommireddy, Anastasya Birger, Bikash Shakya, Allie Crawley, Matthew Nethery, Chuck Pepe-Ranney, Hui-Chia Yu-Kemp, Andy Chan, Tim Schwochert, Drew Kelso, Logan Brown, Julia Portocarrero, Lucas Ribeiro, Victor Bartsevich, Salem Faham, David Wiley, John Russell, Sarah Compton, Joel Parker, Philip Borden, Michael Coyle, Ron Chong, Tedd Elich
ElevateBio Life Edit, Durham, North Carolina, USA



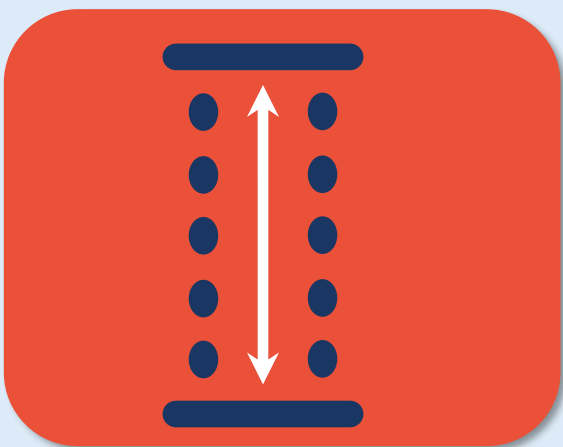
Background

Extensive portfolio of validated CRISPR systems

- Novel RNA-guided CRISPR editors validated both *in vitro* and *in vivo*
- Compact systems fit for both AAV and LNP delivery
- Large collection of natural and engineered PAMs for full access of disease loci



DEPTH AND BREADTH OF GENE EDITING TECHNOLOGIES

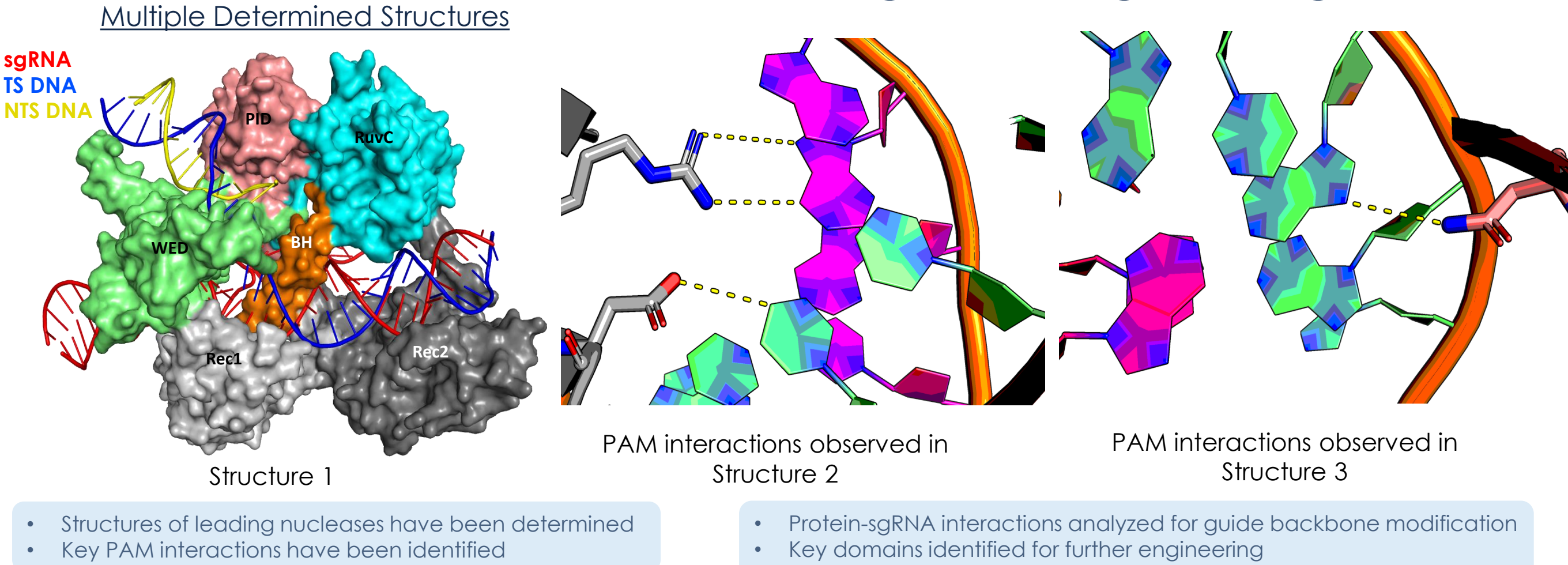


Comprehensive editing modalities

- Knock-out
- A** and **C** base editing
- Gene writing with RT editing
- Expanding to additional modalities

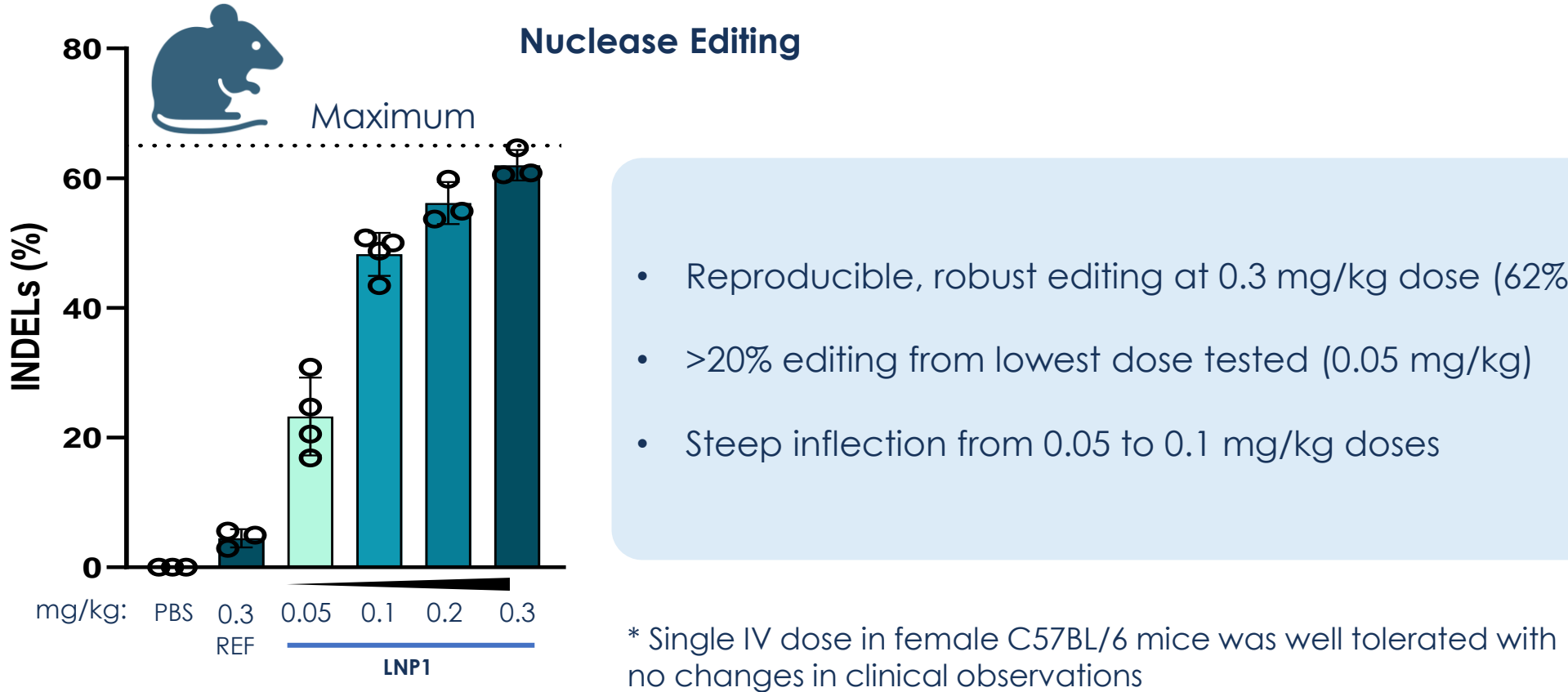
PAM engineering increases access to potential therapeutic targets

Cryo-EM structure of lead proteins guides engineering efforts

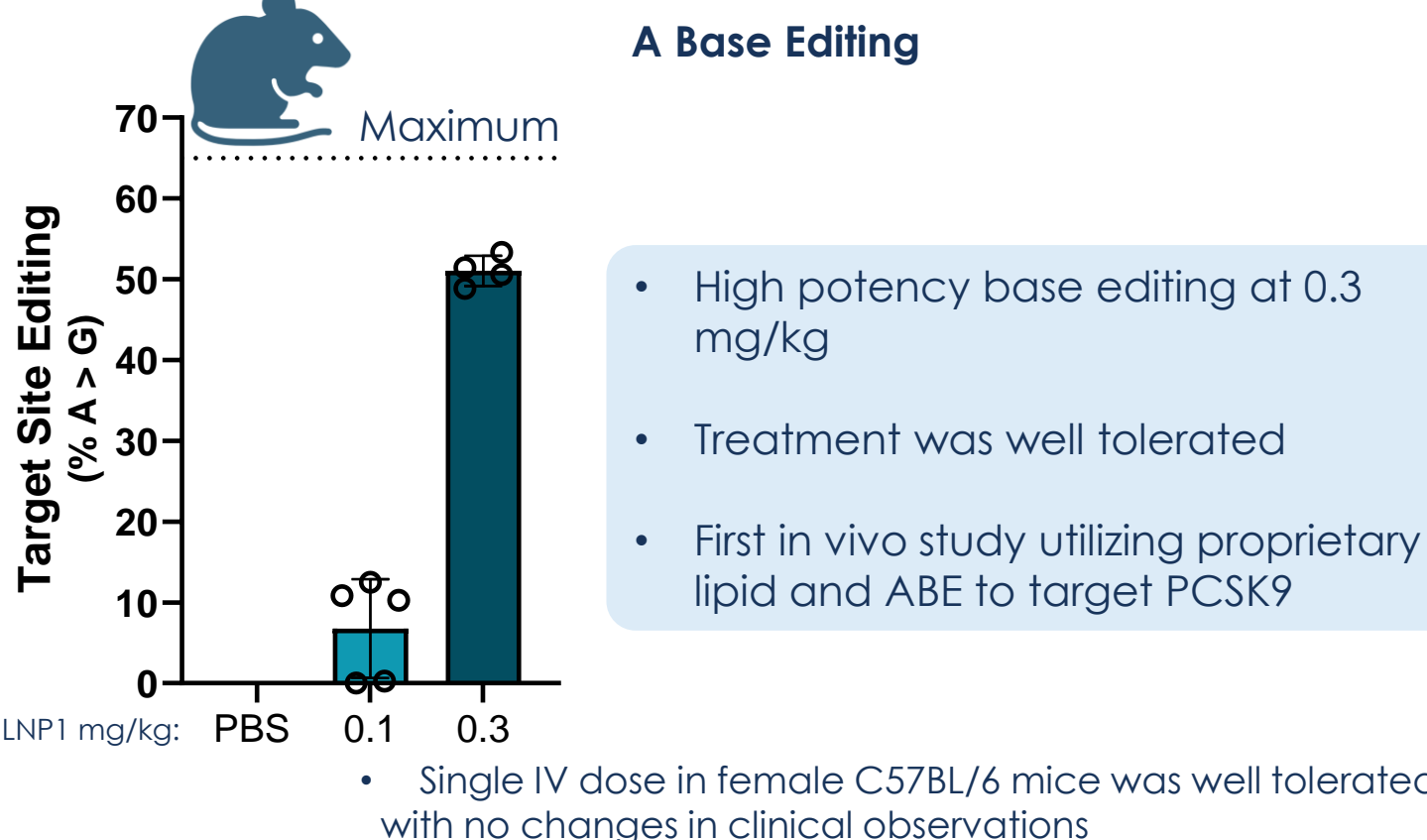


Potent *in vivo* editing using proprietary LNPs for nuclease, base and RT editors

In vivo editing of *Hao1* with Life Edit LNPs

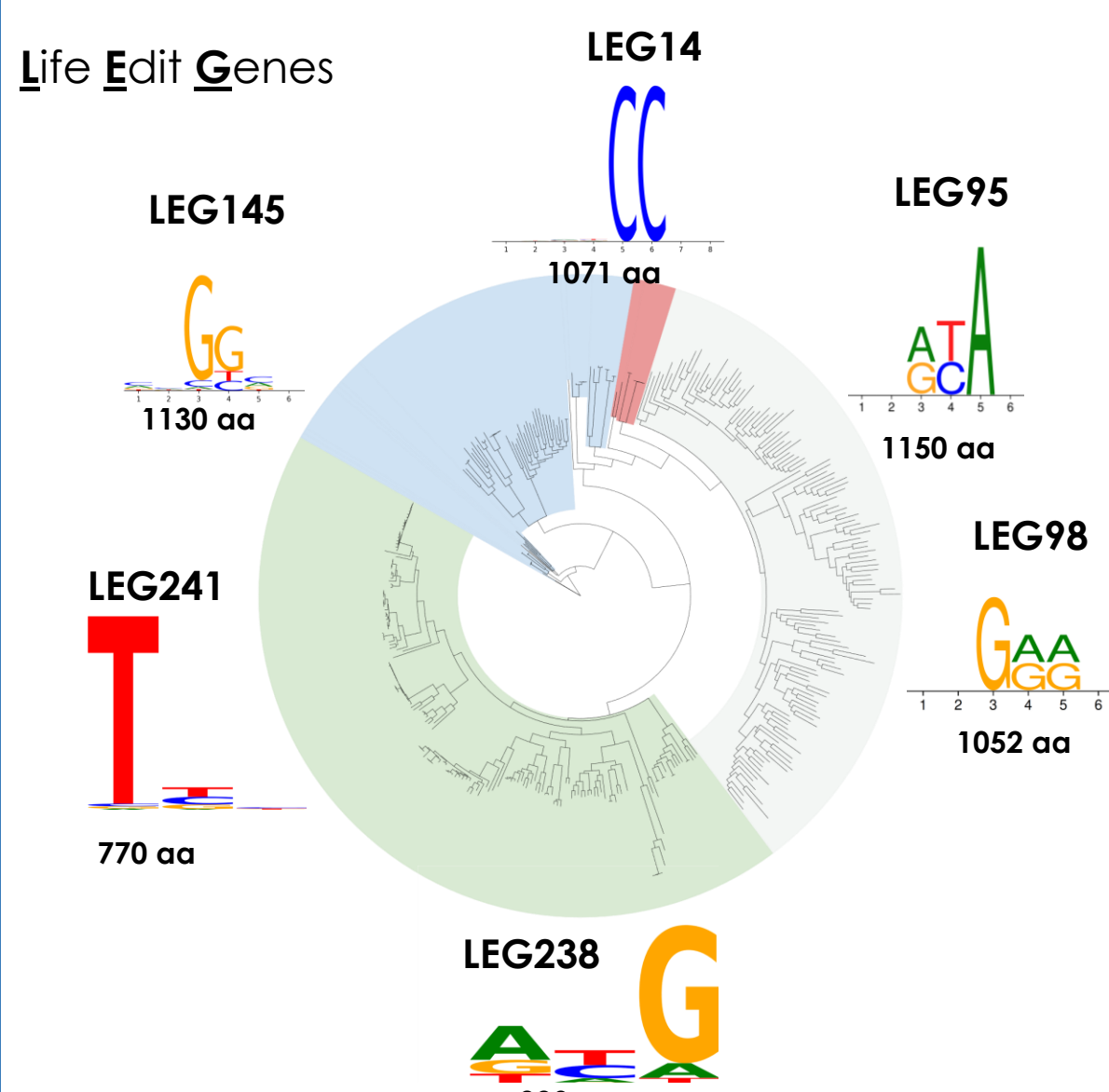


In vivo *Pcsk9* base editing

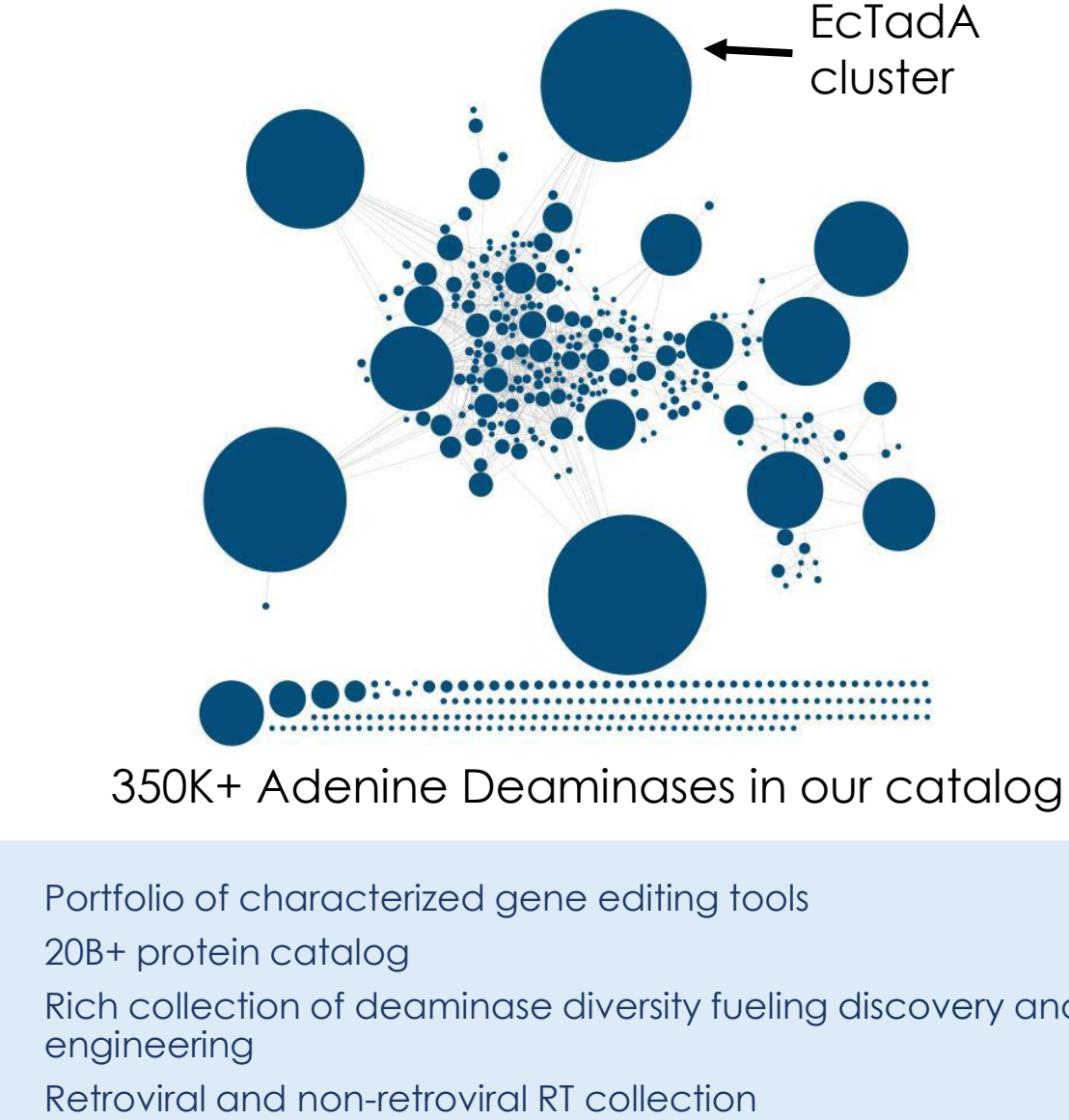


Robust discovery engine allows more options for editing

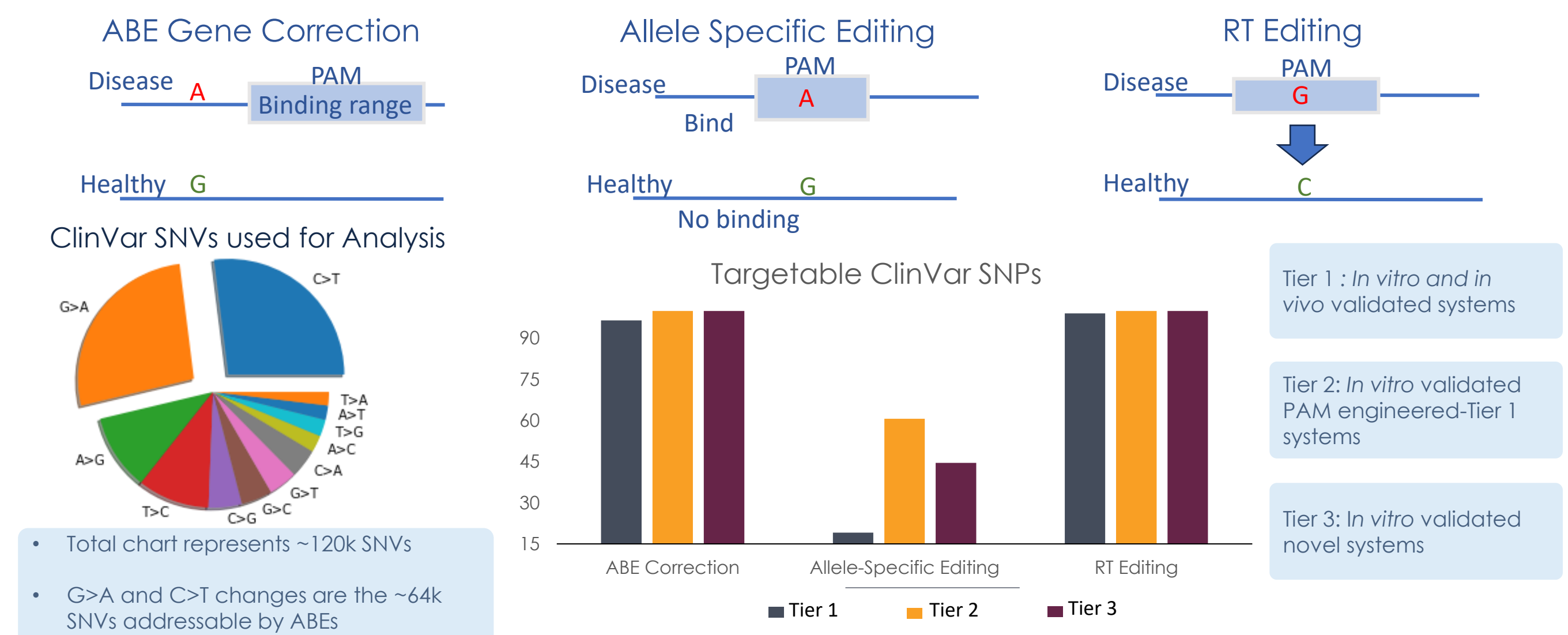
Type II and Type V collection



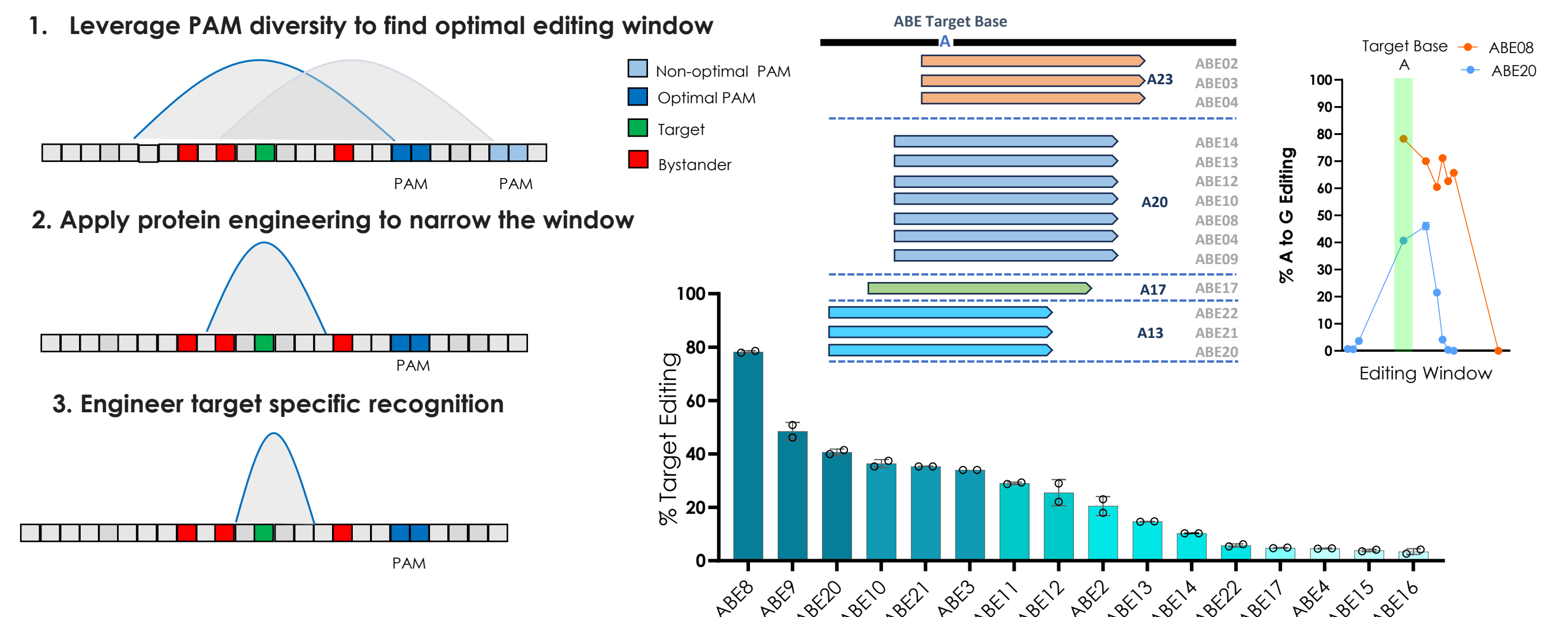
Deaminase and RT collection



CRISPR portfolio targeting >94% ClinVar disease mutations



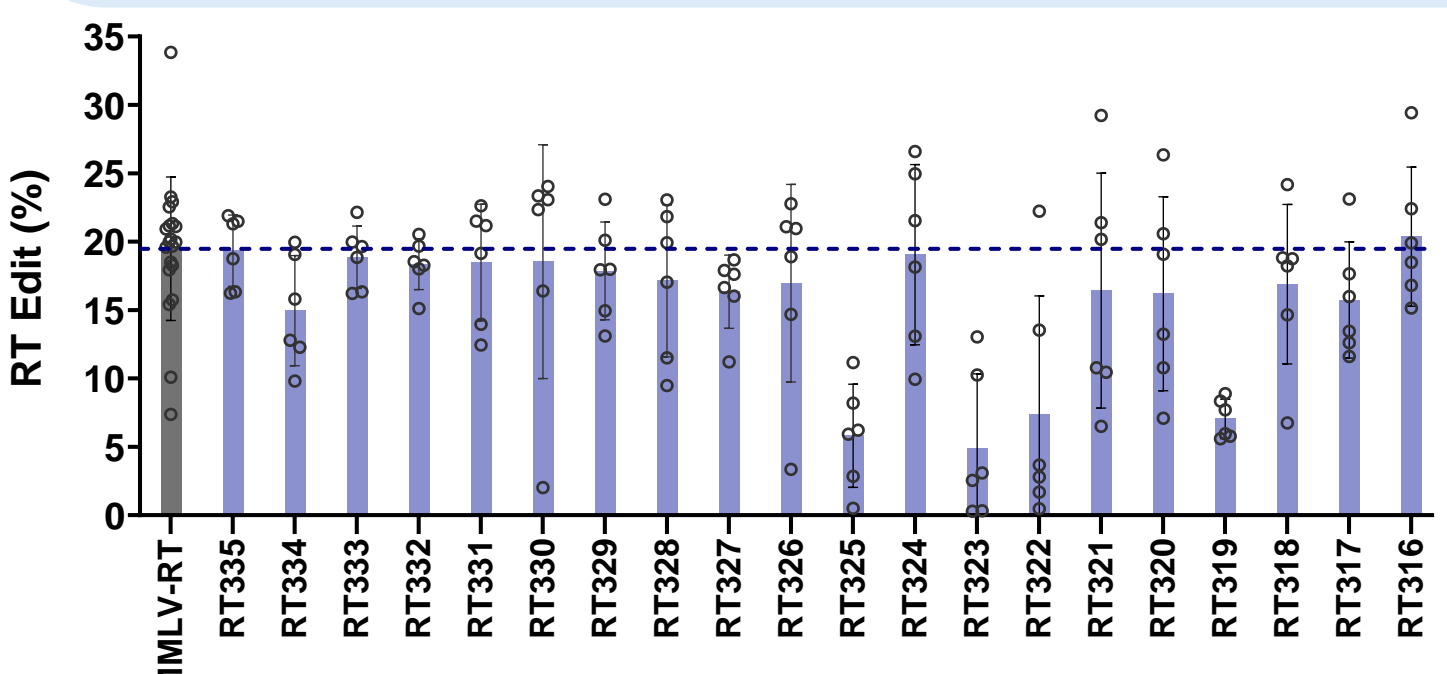
PAM diversity enables optimal positioning of target edit



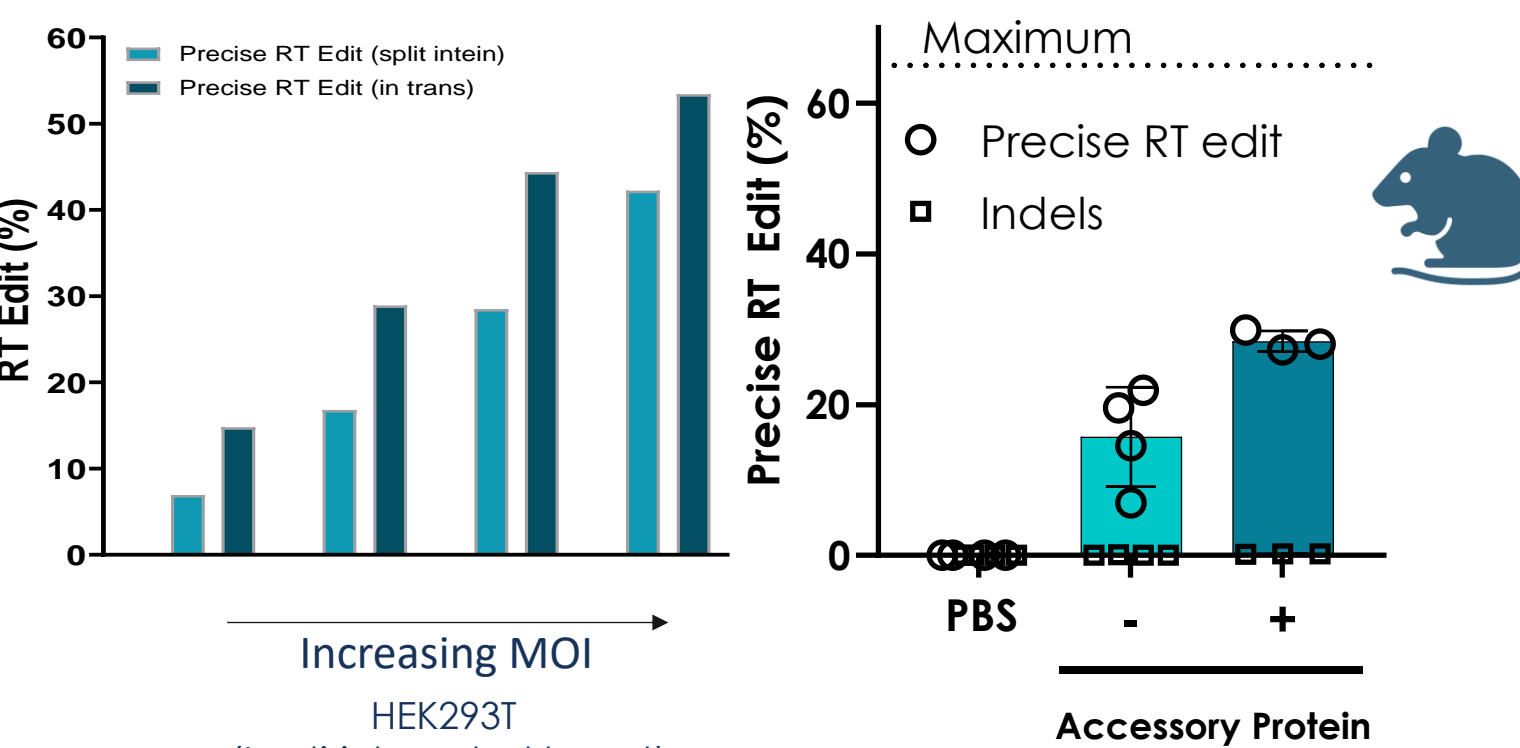
In vitro and *in vivo* demonstration of reverse transcriptase (RT) editing

Life Edit RT Platform:

- Utilizes proprietary LEGs in their nickase form
- Rewrite DNA with precise insertions, deletions and substitutions
- No DNA double-strand breaks required
- Developing Dual RT editing strategies
- Multiple retroviral and non-retroviral RT systems that perform similar to or better than MMLV-RT system



Dual AAV Delivery *in vitro*



LNP-RT Editor targeting in Mouse Liver

- 50% of the maximum possible RT editing in liver
- Minimal/no INDELs observed

Conclusions

- We have developed a large portfolio of compact RNA-guided nucleases with mammalian activity and with diverse PAM recognition allowing full genome access of human disease mutations
- We have validated, both *in vitro* and *in vivo*, a wide range of editing modalities, including RNA guided nucleases, base and RT editors, with multiple delivery methods
- We leverage AI/machine learning, structure guided rational design, automated HT screening and directed evolution to customize our editors for target-specific potency and specificity
- Our proprietary LNPs and AAV vectors plus ex vivo capabilities offer flexibility in delivery of genomic medicines to different organs and tissues
- Our technology platforms accelerate identification of initial hits and developing them into viable clinical candidates

Learn more



vkommireddy@lifeeditinc.com

